Title

In silico Studies of Protein-DNA Interaction & Aggregation of Disease-related Proteins

Abstract

In contrast to the naive expectation based on stoichiometry, recent single-molecule studies on the dynamics of DNA-binding proteins have challenged the standard thermodynamic model of gene regulation. The standard model of protein dissociation from DNA, based on first-order unimolecular reaction scheme, fails under many *in vitro* and *in vivo* contexts. Specifically, the dissociation process is accelerated by increasing the concentration of the protein. However, the community has been puzzled by the molecular mechanism underlying this facilitated dissociation. In this study, I use the state-of-the-art coarse-grained protein-DNA force field to explore the binding landscapes of Fis protein (one nucleoid-associated protein in *E. coli.*) dissociating from DNA. The result provides a unique structural insight into the formation of Fis intermediate on DNA. When two Fis molecules are present, simulations uncover a ternary complex, where the originally bound Fis protein is partially dissociated from DNA. The simulations support a three-state sequential kinetic model for facilitated dissociation, thus explaining concentration-dependent dissociation.

Although aggregation of Amyloid β (A β) has received much attention among protein scientists, the question regarding why A β 42 aggregates faster than A β 40 is still one major puzzle in the community. Due to the transient nature of A β oligomers and small aggregates, the key to the question apparently lies in the nucleation at the early stages of A β aggregation. We solve this puzzle by exploring and comparing the aggregation free energy landscapes of A β 40 and A β 42 which allow us to accurately capture the key molecular event, backtracking, that governs the prefibrillar-to-fibrillar conformational conversion of A β 40. The backtracking is responsible for slowing the aggregation process. This computational aggregation protocol along with the taxonomy of oligomer clusters that I develop offer a theoretical framework that forms the basis for a library of fully atomistic oligomer structures which may offer specific targets for drug design.